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(21) International Application Number: PCT/US99/19990 (22) International Filing Date: 31 August 1999 (31.08.99) (30) Priority Data: 60/098,689 31 August 1998 (31.08.98) US (71) Applicant: OREGON HEALTH SCIENCE UNIVERSITY [US/US]; 3181 Southwest Sam Jackson Park Road, Portland, OR 97201-3098 (US). (72) Inventors: NELSON, Jay; 21067 S.W. Meadow Way, Tualatin, OR 97062 (US). STREBLOW, Daniel; 12508 S.W. North Dakota Street, Tigard, OR 97223 (US). SODERBERG-NAUDER, Cecelia; Karolinska Institute, S-Stockholm (SE). SMITH, Patricia; 945 N.E. Hazelfern Place, Portland, OR 97232 (US). RUCHTI, Franziska; 3735 S.W. Bridlemile Lane, Portland, OR 97221 (US). (74) Agent: OSTER, Jeffrey, B.; Davis Wright Tremaine LLP, 2600 Century Square, 1501 4th Avenue, Seattle, WA 98101-1688 (US).	(81) Designated States: AU, CA, JP, Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM). Published <i>With international search report. Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.</i>	
(54) Title: PREVENTION OF CELL MIGRATION INITIATION WITH CMV US28 RECEPTOR ANTAGONISTS (57) Abstract There is disclosed an assay system for determining therapeutic activity for treating restenosis, atherosclerosis, chronic rejection syndrome and graft versus host disease (GVHD) by measuring inhibition of cell migration activity in smooth muscle cells expressing a US28 receptor from the CMV genome. Specifically, there is disclosed a method for measuring inhibition of cell migration in isolated cells transfected with US28 or infected with CMV and stimulated with a ligand. There is further disclosed a method for treating atherosclerosis, restenosis, chronic rejection syndrome and graft versus host disease (GVHD), comprising administering an effective amount of an agent that is a US28 receptor antagonist, wherein a US28 receptor antagonist comprises an inhibitor compound that prevents transduction of US28 receptor signal stimulated by a US28 receptor ligand, wherein a US28 receptor ligand is selected from the group consisting of RANTES, MIP-1 α and MCP. The invention further provides a method for treating restenosis, atherosclerosis, chronic rejection syndrome and GVHD by administering KHSV encoded vMIP-2, fractalkine or herbimycin.		

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PREVENTION OF CELL MIGRATION INITIATION WITH CMV US28 RECEPTOR ANTAGONISTS

Technical Field of the Invention

5 The present invention provides an assay system for determining therapeutic activity for treating restenosis, atherosclerosis, chronic rejection syndrome and graft versus host disease (GVHD) by measuring inhibition of cell migration activity in smooth muscle cells expressing a US28 receptor from the CMV genome. Specifically, the present invention provides a method for measuring inhibition of cell migration in isolated cells transfected with US28 or infected
10 with CMV and stimulated with a ligand. The invention further provides a method for treating restenosis, atherosclerosis, chronic rejection syndrome and GVHD by administering KHSV encoded vMIP-2, fractalkine or herbimycin.

Background of the Invention

15 Atherosclerosis, Restenosis, Chronic Rejection Syndrome and GVHD

Atherosclerosis is a major cause of morbidity in the industrialized world. Atherosclerotic lesions usually become apparent in adult patients as a result of complete occlusion of a strategic blood vessel and the resulting complication. However, such lesions begin much earlier in the life of the patient. It was later noticed that there was a statistical
20 association with viral infection, particularly CMV.

It has been postulated that CMV and possibly herpes virus are involved in the inducement of atherosclerotic lesions. Several investigators have demonstrated the presence of CMV nucleic acids and/or antigens in the human arterial wall using DNA hybridization techniques (Melnick et al., *Lancet* 11:644-647, 1983), immunohistochemistry (Petrie et al., *J.*
25 *Infect. Dis.* 155:158-159, 1987), dot blot and *in situ* hybridization techniques (Hendrix et al., *Am. J. Path.* 134:1151-1157, 1989), and by polymerase chain reaction (PCR) techniques using probes derived from immediate early and late genomic regions (Hendrix et al., *Am. J. Path.* 136:23-28, 1990). Thus, there has been finding of viral antigens and nucleic acid sequences in arterial smooth muscle cells that suggest that CMV infection of the arterial wall may be a
30 common occurrence in patients with atherosclerosis.

Soon after renal transplantation became an accepted treatment, an association was noted between CMV infection, glomerulopathy, and rejection of the transplanted kidney. Thus, CMV was investigated to determine if it played a role in graft atherosclerosis that frequently occurs after heart transplantation (Grattan et al., *J. Am. Med. Assn.* 261:3561-3566, 1989). The findings
35 show that heart transplant patients who are immunosuppressed and become infected with CMV are particularly prone to develop atherosclerosis in the transplanted organ. It is postulated that the artery wall may be the site of CMV latency because CMV DNA but not infectious virus was found in the artery wall.

Role of Chemokines

Chemokines are chemoattractants for neutrophils, monocytes, lymphocytes and bone marrow progenitors, as well as other cell types. The family of chemokines comprises four subfamilies, defined by the distribution of cysteine residues in the N terminus of these factors, the CXC, CC, C, and CX3C subfamilies. The chemokines are related by primary structure, particularly by conservation of a four-cysteine motif. C-C chemokines include such members as human monocyte chemotactic protein 1 (MCP-1), RANTES, and the macrophage inflammatory proteins 1 α and 1 β (MIP-1 α and MIP-1 β). These ligands exhibit chemoattractant potential for monocytes but not neutrophils. CMV infection can also modify the level of chemokines. The level of RANTES (a chemokine) produced by cells recovered by bronchoalveolar lavage from lung transplant patients with CMV pneumonitis shows that cells from infected patients secreted greater amounts of RANTES than did cells recovered from either patients undergoing acute rejection or from control subjects (Monti et al., *Transplantation* 61:1757-1762, 1996). AIDS patients with CMV encephalitis have higher concentrations of MCP-1 (a chemokine) but not other chemokines in their spinal fluid than do HIV seropositive persons who are asymptomatic or AIDS patients with a number of other opportunistic infections of the central nervous system (Bernasconi et al., *J. Infec. Dis.* 174:1098-1101, 1996). When fibroblasts were infected with CMV, RANTES mRNA and protein expression are induced early, but extracellular RANTES accumulation, but not transcription is down-regulated late during CMV infection (Michelson et al., *J. Virol.* 71:6495-6500, 1997). Therefore, CMV infection has the capacity to both induce cell migration and enhance chemokine production early during the infection process.

Chemokine receptors tend to be multiple membrane-spanning proteins, generally 7 or 8 membrane-spanning proteins and tend to transduce signal through G-coupled protein signal transduction. Human C-C chemokines tend to bind to the US28 receptor of CMV (Neote et al., *Cell* 72:415-425, 1993). There is also a sequence homology between the C-CKR-1 receptor (normal human gene) and the CMV US28 sequence in the open reading frame region. (Neote et al., 1993). Thus, Neote et al. speculated that "the protein encoded by the US28 open reading frame of Towne strain CMV can bind C-C chemokines but not the C-X-C chemokine IL-8. However, none of the earlier chemokine receptor papers, including Neote et al., has made the connection between US28 and its role in mediating smooth muscle cell proliferation.

Summary of the Invention

The present invention provides an assay for determining therapeutic activity of US28 receptor antagonists, comprising (a) obtaining and isolating smooth muscle cells into a first chamber of a migration device, wherein the first migration chamber comprises growth media chambers and is defined by a first side of a membrane and chamber walls, and wherein the migration device comprises a second chamber defined by the second side of the membrane and having an enclosed space; (b) infecting the smooth muscle cells with human cytomegalovirus (HCVM) containing a gene encoding the US28 receptor; (c) adding a candidate therapeutic agent to the first chamber; and (d) determining the amount of cellular migration into the second chamber, whereby inhibition of cellular migration of infected smooth muscle cells indicates

therapeutic activity. Preferably, the smooth muscle cells are isolated from pulmonary arteries. Preferably, the membrane has a pore size of from about 2 to about 10 microns. Most preferably, the membrane pore size is about 3 microns. Preferably the amount of cellular migration is determined by an assay for counting the number of smooth muscle cells in the second chamber wherein the assay for counting the number of smooth muscle cells is selected from the group consisting of microscopic cell counting per unit area, radiolabeling the smooth muscle cells and counting radioactivity in the second chamber, attaching a fluorescent probe to the smooth muscle cells and measuring fluorescence within the second chamber, and combinations thereof.

The present invention further provides a method for treating atherosclerosis, restenosis, chronic rejection syndrome and graft versus host disease (GVHD), comprising administering an effective amount of an agent that is a US28 receptor antagonist, wherein a US28 receptor antagonist comprises an inhibitor compound that prevents transduction of US28 receptor signal stimulated by a US28 receptor ligand, wherein a US28 receptor ligand is selected from the group consisting of RANTES, MIP-1 α and MCP. Preferably, the US28 receptor antagonist is selected from the group consisting of an antibody that binds to an extracellular portion of the US28 receptor, and an antisense oligonucleotide having a nucleic acid sequence antisense to the US28 cDNA and inhibiting translation of US28 expression in infected smooth muscle cells, or a US28 binding antagonist, wherein the US28 binding antagonist is selected from the group consisting of KHSV encoded vMIP-2, fractalkine, and herbimycin. Preferably, the monoclonal antibody is chimeric or humanized by means for humanizing non-human antibodies. Preferably, the US28 antisense sequences are selected from the group consisting of SEQ ID NOS. 2-28.

The present invention further provides a method for enhancing cellular migration, comprising infecting a cell with a viral nucleic acid containing a gene encoding CVM US28 receptor or transfecting a cell with a vector comprising the cDNA sequence for US28 operably linked to a viral promoter sequence, and stimulating the transfected or infected cell with a US28 receptor ligand, selected from the group consisting of RANTES, MIP-1 α and MCP1.

Brief Description of the Drawings

Figure 1 shows the ability of HCMV-GFP (human cytomegalovirus GFP) to infect pulmonary smooth muscle cells (SMC) *in vitro*. SMC were infected with HCMV-GFP (MOI 10) for 2 days and then examined for the presence of GFOHCMV immediate early expression. Colocalization of GFP and IE (intermediate-early) were observed only in HCMV-GFP infected cells.

Figure 2 shows that HCMV infected SMCs were examined by electron microscopy for the presence of virus. The photos show numerous virus capsids were found in the nucleus and mature virions were observed on the plasma membrane of HCMV infected cells.

Figure 3 shows a one-step growth curve showing HCMV replication in SMCs. SMC were infected with HCMV Towne strain at MOI1. These data show HCMV growth and release in SMC exhibited normal kinetics.

Figure 4 shows that HCMV infection induced actin reorganization in SMC as an

indication of migration activity. The cells were treated with either PDGF (100 ng/ml) of infected with HCMV Towne strain at MOI1. Actin distribution was visualized at 5 days post-treatment by fluorescence using TRITC conjugated phalloidin.

Figure 5 shows an SMC migration assay scheme. SMCs are cultured in the upper chamber and infected with HCMV (preferably at or near MOI1). Only cells that are infected will migrate through a filter (preferably 3 micron) to the lower chamber. The cells in the lower chamber are counted by microscopy or labeled with a radioactive or fluorescent label.

Figure 6 shows the presence of HCMV in migrating SMCs. SMC were infected with HCMV-GFP. The migrating cells were analyzed for the presence of GFP and HCMV glycoprotein gB by immunofluorescence using anti-gB antibodies. All of the migrating SMCs exhibited GFP and gB expression.

Figure 7 shows that HCMV induced migration of SMCs. Cellular mobility assays were used to determine the specificity of HCMV induced SMC migration. SMCs that were treated with PDGF (Platelet-derived growth factor, an inducer of cellular migration) did not cause cell migration in SMCs to nearly the extent as HCMV infection. Moreover, HCMV neutralizing antibodies reduced cellular migration to mock levels.

Figure 8 shows that HCMV-induced migration was specific for SMCs. Both SMCs and human foreskin fibroblasts (HFF) were infected and analyzed in migration assays. The data provided in Figure 8 show that HCMV induced cellular migration occurred only in SMCs but not in similarly-infected HFFs.

Figure 9 shows that protein synthesis was required for HCMV-induced SMC migration. SMC mobility was blocked by cyclohexamide (a general protein synthesis inhibitor). Moreover, foscarnet (an inhibitor of HCMV late gene production) did not inhibit SMC migration.

Figure 10 shows that HCMV genome encodes four putative chemokine receptors, including US27, US28, UL33 and UL78.

Figure 11 shows that HCMV infection of HFFs induced RANTES (chemokine) expression. HFF cell culture supernatants were collected every 8 hours from infected HFFs and RANTES concentrations in the supernatants were determined by an ELISA assay.

Figure 12 shows that the addition of RANTES at the concentrations shown to HCMV-GFP-infected SMCs increased SMC migration in a dose-dependent manner.

Figure 13 shows that that a HCMV having the US28 receptor gene deleted affected cell motility.

Figure 14 shows the construction scheme for human CMV GFP recombinants.

Figure 15 shows the results of inhibition of a PTK pathway effect of US28 SMC migration with several PTK inhibitors including herimycin A, pertussis toxin and genistein at the concentrations indicated. Pertussis toxin had no effect.

Detailed Description of the Invention

US28 Receptor

The sequence characterization of the US28 receptor is provided in Neote et al. (*Cell*

72:415-425, 1993) and also the cDNA sequence is SEQ ID No. 1. The present invention is based upon the discovery that the CMV effect in causing smooth muscle proliferation and an initiating event in the diseases atherosclerosis, restenosis, chronic organ rejection and GVHD, is mediated primarily through signal transduction in infected smooth muscle cells through the US28 receptor. Based upon this discovery, described herein, the claimed invention is provided that provides US28 receptor antagonist molecules that have therapeutic effect. Moreover, the present invention provides an assay procedure to screen of other US28 antagonist molecules that, based upon the findings reported here, are effective for treating atherosclerosis, restenosis, chronic organ rejection and GVHD.

Role of US28 in Smooth Muscle Cell Migration

The present invention is based upon the discovery of the role of US28 in mediating the properties of CMV to stimulate smooth muscle cells that can ultimately lead to atherosclerosis, restenosis, chronic rejection syndrome or GVHD in susceptible patients. It is clear to a skill practitioner that a patient must first have been a transplant recipient before he or she is at risk for either GVHD or chronic rejection syndrome. Moreover, restenosis first requires an angioplasty-type procedure or other chemical or surgical intervention in clearing occluded or partially occluded arteries before the patient is at risk for restenosis.

The data provided in the figures (1-13) show that CMV virus infects smooth muscle cells *in vitro* and that intermediate early expression of viral protein can be seen (Figures 1-3). The affect of CMV infection in smooth muscle cells is shown affect actin reorganization as a marker for migration activity (Figure 4). Moreover, the infected cells were able to migrate in a migration chamber, such as the one shown in Figure 6. The scheme shown in Figure 5 provides that only infected cells have the capability to migrate through a filter in a migration chamber. Thus, smooth muscle cells infected with CMV showed the ability to migrate, even to a much greater extent than non-infected smooth muscle cells treated with the migration enhancing growth factor, PDGF (Figure 7).

The next set of experiments were designed to determine which protein or proteins, encoded by the VMC genome, was responsible for conferring the migration activity on infected smooth muscle cells. It was first found that protein synthesis was required to confer the migration activity on infected smooth muscle cells (Figure 9). Moreover, the suspect protein or proteins encoded by the CMV genome were not late gene production genes as evidenced by the fact that foscarnet (an inhibitor of HCMV late gene production) did not inhibit migration of infected smooth muscle cells (Figure 9). This left four putative chemokine receptors that are encoded by the CMV genome, US27, US28, UL33 and UL78 (Figure 10). In knock-out experiments, wherein each of the four foregoing chemokine receptor genes were knocked out, it was only a US28 knock out that was able to inhibit smooth muscle cell migration activity when smooth muscle cells were infected with the US28 knock out variety of CMV (Figure 13). However, the ability of CMV infection to increase cell migration of smooth muscle cells may not be only as US28 affect and there are also ligand activity that needs to activate the US28

receptor. Moreover, CMV infection seems to also have an autocrine function in enhancing certain C-C chemokine production, such as RANTES (Figures 11-12). Accordingly, the foregoing data provides the basis for the present invention.

Screening Assay

5 The present invention provides an assay for determining therapeutic activity of US28 receptor antagonists, comprising (a) obtaining and isolating smooth muscle cells into a first chamber of a migration device, wherein the first migration chamber comprises growth media chambers and is defined by a first side of a membrane and chamber walls, and wherein the migration device comprises a second chamber defined by the second side of the membrane and
10 having an enclosed space; (b) infecting the smooth muscle cells with human cytomegalovirus (HCMV) containing a gene encoding the US28 receptor; (c) adding a candidate therapeutic agent to the first chamber; and (d) determining the amount of cellular migration into the second chamber, whereby inhibition of cellular migration of infected smooth muscle cells indicates therapeutic activity. Preferably, the smooth muscle cells are isolated from pulmonary arteries.
15 Preferably, the membrane has a pore size of from about 2 to about 10 microns. Most preferably, the membrane pore size is about 3 microns. Preferably the amount of cellular migration is determined by an assay for counting the number of smooth muscle cells in the second chamber wherein the assay for counting the number of smooth muscle cells is selected from the group consisting of microscopic cell counting per unit area, radiolabeling the smooth muscle cells and
20 counting radioactivity in the second chamber, attaching a fluorescent probe to the smooth muscle cells and measuring fluorescence within the second chamber, and combinations thereof. Preferably, the infected smooth muscle cells are further stimulated with ligand to enhance migration activity, wherein the ligand is a C-C chemokine. Preferably, the C-C ligand is selected from the group consisting of RANTES, MCP-1, MIP-1 α , MIP-1 β , and combinations
25 thereof.

KHSV-Encoded vMIP-2

 KSHV-(Kaposi's sarcoma-associated herpes virus) encoded vMip alpha and beta has been described as having angiogenic and HIV inhibitory functions (Boshoff et al., *Science* 278:290-294, 1997). It has also been described as a broad-spectrum chemokine antagonist
30 (Kledal et al., *Science* 277:1656-1659, 1997). The present invention adds to the therapeutic uses for KHSV-encoded MIP for treating atherosclerosis, restenosis, chronic rejection syndrome and GVHD.

Fractalkine

 Results from several studies showed that the polypeptide fractalkine is a ligand for US28
35 receptor and functions as a US28 antagonist through competitive binding. Fractalkine has been described in Kledal et al. *FEBS Lett.* 441:209-214, 1998. The present invention adds to the therapeutic uses for fractalkine for treating atherosclerosis, restenosis, chronic rejection syndrome and GVHD.

Herbimycin

Herbimycin A is a PTK (protein tyrosine kinase) pathway inhibitor. It is available commercially (Sigma). In Figure 15, the effect of herbimycin A on US28 SMC (smooth muscle cell) migration showed herbimycin A was effective in inhibiting US28 transfected smooth muscle cell migration. Thus, it appears that US28 SMC migration is mediated through a PTK pathway. The present invention adds to the therapeutic uses for herbimycin for treating atherosclerosis, restenosis, chronic rejection syndrome and GVHD.

Antisense

US28 is made off of two different transcripts, one only contains the US28 ORF and the other contains US27/28 ORF's. Both use the same poly-A signal. Antisense oligo sequences as

US28 antagonists for both US27 and US28 are as follows:

- US27-5'-1---ATT TGT AGA GGT GGT CAT [SEQ ID NO. 9]
- US27-5'-2---GCT CAC CTG CGT TAA GGT [SEQ ID NO. 10]
- US27-5'-3---GTG CTG TTT AAG GTG TGG [SEQ ID NO. 11]
- US27-5'-4---AGT GTA CTC GAA CAA CTG [SEQ ID NO. 12]
- US27-5'-5---CAA CCA TAC CCC GTT GGC [SEQ ID NO. 13]
- US27-3'-1---TTC ACG CAG CAA CAG GCG [SEQ ID NO. 14]
- US27-3'-2---CCT GGT AAG GTA TAT CCT [SEQ ID NO. 15]
- US27-3'-3---GTA GCT CAA TAT CAA TGT [SEQ ID NO. 16]
- US27-3'-4---GCC CTT CTT TGT ATG TCC [SEQ ID NO. 17]
- US27-3'-5---ATG GGT ACG TTT GGT GTG [SEQ ID NO. 18]
- US28-5'-1---CGT CGT CGT CGG TGT CAT [SEQ ID NO. 19]
- US28-5'-2---CGT CGT GAG TTC CGC GGT [SEQ ID NO. 20]
- US28-5'-3---CAG GGA GTC GCT TCA TCG [SEQ ID NO. 21]
- US28-5'-4---TGA TTA AGC ACG TCG GTG [SEQ ID NO. 22]
- US28-5'-5---GAA GAG AAA GAC AAC GCC [SEQ ID NO. 23]
- US28-3'-1---GCT GTG GTA CCA GGA TAC [SEQ ID NO. 24]
- US28-3'-2---CTC CGA CGC GAA AAG CTC [SEQ ID NO. 25]
- US28-3'-3---GTC TCT CTT CGG CTC GGC [SEQ ID NO. 26]
- US28-3'-4---CGG ACA GCG TGT CGG AAG [SEQ ID NO. 27]
- US28-3'-5---GAG ACG CGA CAC GCC TCG [SEQ ID NO. 28]

Additional antisense sequences are provided as SEQ ID NOS 2-8.

Pharmaceutical Formulation

The inventive method in the form of a pharmaceutical composition comprising a US28 antagonist can be administered to a patient either by itself (complex or combination) or in pharmaceutical compositions where it is mixed with suitable carriers and excipients. A US28 antagonist can be administered parenterally, such as by intravenous injection or infusion, intraperitoneal injection, subcutaneous injection, or intramuscular injection. A US28 antagonist can be administered orally or rectally through appropriate formulation with carriers and excipients to form tablets, pills, capsules, liquids, gels, syrups, slurries, suspensions and the like.

A US28 antagonist can be administered topically, such as by skin patch, to achieve consistent systemic levels of active agent. A US28 antagonist is formulated into topical creams, skin or mucosal patch, liquids or gels suitable to topical application to skin or mucosal membrane surfaces. A US28 antagonist can be administered by inhaler to the respiratory tract for local or
5 systemic treatment of HIV infection.

The dosage of the US28 antagonist suitable for use with the present invention can be determined by those skilled in the art from this disclosure. The US28 antagonist will contain an effective dosage (depending upon the route of administration and pharmacokinetics of the active agent) of the US28 antagonist and suitable pharmaceutical carriers and excipients, which are
10 suitable for the particular route of administration of the formulation (*i.e.*, oral, parenteral, topical or by inhalation). The active US28 antagonist is mixed into the pharmaceutical formulation by means of mixing, dissolving, granulating, dragee-making, emulsifying, encapsulating, entrapping or lyophilizing processes. The pharmaceutical formulations for parenteral administration include aqueous solutions of the active US28 antagonist in water-soluble form.
15 Additionally, suspensions of the active US28 antagonist may be prepared as oily injection suspensions. Suitable lipophilic solvents or vehicles include fatty oils such as sesame oil, or synthetic fatty acid esters, such as ethyl oleate or triglycerides, or liposomes. Aqueous injection suspensions may contain substances, which increase the viscosity of the suspension, such as sodium carboxymethyl cellulose, sorbitol, or dextran. The suspension may optionally contain
20 stabilizers or agents to increase the solubility of the complex or combination to allow for more concentrated solutions.

Pharmaceutical formulations for oral administration can be obtained by combining the active compound with solid excipients, such as sugars (*e.g.*, lactose, sucrose, mannitol or sorbitol), cellulose preparations (*e.g.*, starch, methyl cellulose, hydroxypropylmethyl cellulose,
25 and sodium carboxymethyl cellulose), gelatin, gums, or polyvinylpyrrolidone. In addition, a disintegrating agent may be added, and a stabilizer may be added.

SEQUENCE LISTING

(1) GENERAL INFORMATION:

30 (i) APPLICANTS: Nelson, Jay et al.

(ii) TITLE OF INVENTION: Prevention of Cell Migration
Initiation with CMV US28 Receptor Antagonists

35 (iii) NUMBER OF SEQUENCES: 8

(iv) CORRESPONDENCE ADDRESS:

(A) ADDRESSEE: DAVIS WRIGHT TREMAINE

(B) STREET: 1501 Fourth Avenue, 2600 Century Square

40 (C) CITY: Seattle

(D) STATE: Washington

(E) COUNTRY: U.S.A.

(F) ZIP: 98101

(v) COMPUTER READABLE FORM:

- (A) MEDIUM TYPE: Floppy disk
 (B) COMPUTER: PC compatible
 (C) OPERATING SYSTEM: Windows95
 (D) SOFTWARE: Word

(vi) CURRENT APPLICATION DATA:

- (A) APPLICATION NUMBER: to be assigned
 (B) FILING DATE: 31 August 1999
 (C) CLASSIFICATION:

(viii) ATTORNEY/AGENT INFORMATION:

- (A) NAME: Oster, Jeffrey B.
 (B) REGISTRATION NUMBER: 32,585
 (C) REFERENCE/DOCKET NUMBER:

(ix) TELECOMMUNICATION INFORMATION:

- (A) TELEPHONE: 206 628 7711
 (B) TELEFAX: 206 628 7699

(2) INFORMATION FOR SEQ ID NO:1:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1087
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: US28 Receptor coding region

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

30	1	AAACGTCATC	TCGCCGACGT	GGTGAACCGC	TCATATAGAC	CAAACCGGAC	50
	51	GCTGCCTCAG	TCTCTCGGTG	CGTGGACCAG	ACGGCGTCCA	TGCACCGAGG	100
	101	GCAGAACTGG	TGCTATCATG	ACACCGACGA	CGACGACCGC	GGAACCTCAG	150
	151	ACGGAGTTTG	ACTACGATGA	AGACGCGACT	CCTTGTGTTT	TCACCGACGT	200
	201	GCTTAATCAG	TCAAAGCCAG	TTACGTTGTT	TCTGTACGGC	GTTGTCTTTC	250
35	251	TCTTCGGTTC	CATCGGCAAC	TTCTTGGTGA	TCTTCACCAT	CACCTGGCGA	300
	301	CGTCGGATTG	AATGCTCCGG	CGATGTTTAC	TTTATCAACC	TCGCGGCCGC	350
	351	CGATTTGCTT	TTCGTTTGTA	CACTACCTCT	GTGGATGCAA	TACCTCCTAG	400
	401	ATCACAACTC	CCTAGCCAGC	GTGCCGTGTA	CGTTACTCAC	TGCCTGTTTC	450
	451	TACGTGGCTA	TGTTTGCCAG	TTTGTGTTTT	ATCACGGAGA	TTGCACTCGA	500
40	501	TCGCTACTAC	GCTATTGTTT	ACATGAGATA	TCGGCCTGTA	AAACAGGCCT	550
	551	GCCTTTTCAG	TATTTTTTGG	TGGATCTTTG	CCGTGATCAT	CGCCATTCCA	600
	601	CACTTTATGG	TGGTGACCAA	AAAAGACAAT	CAATGTATGA	CCGACTACGA	650
	651	CTACTTAGAG	GTCAGTTACC	CGATCATCCT	CAACGTAGAA	CTCATGCTTG	700
	701	GTGCTTTTCG	GATCCCGCTC	AGTGTTATCA	GCTACTGCTA	CTACCGCATT	750
45	751	TCCAGAATCG	TTGCGGTGTC	TCAGTCGCGC	CACAAAGGTC	GCATTGTACG	800
	801	GGTACTTATA	GCGGTCGTGC	TTGTCTTTAT	CATCTTTTGG	CTGCCGTACC	850
	851	ACCTAACGCT	GTTTGTGGAC	ACGTTAAAAC	TCCTCAAATG	GATCTCCAGC	900
	901	AGCTGCGAGT	TCGAAAGATC	GCTCAAACGT	GCGCTCATCT	TGACCGAGTC	950
	951	GCTCGCCTTT	TGTCAGTGTT	GTCTCAATCC	GCTGCTGTAC	GTCTTCGTGG	1000
50	1001	GCACCAAGTT	TCGGCAAGAA	CTACACTGTC	TGCTGGCCGA	GTTTCGCCAG	1050
	1051	CGACTCTTTT	CCGCGATGT	ATCCTGGTAC	CACAGCA	1087	

(2) INFORMATION FOR SEQ ID NO:2:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: US28 receptor antisense

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

10 CGGAATTAGTC AGTTTCGGTC 20

(2) INFORMATION FOR SEQ ID NO:3:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: US28 receptor antisense

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

20 CGTCTTGACC ACGATAGTAC 20

(2) INFORMATION FOR SEQ ID NO:4:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: US28 receptor antisense

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

30 GCAGCCTAAG TTACGAGGCC 20

(2) INFORMATION FOR SEQ ID NO:5:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: US28 receptor antisense

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

40 TAGTGTTGAG GGATCGGTCTG 20

(2) INFORMATION FOR SEQ ID NO:6:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: US28 receptor antisense

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

50 CGGAATTAGTC AGTTTCGGTC 20

- (2) INFORMATION FOR SEQ ID NO:7:
 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 20
 (B) TYPE: nucleic acid
5 (C) STRANDEDNESS: single
 (D) TOPOLOGY: unknown
 (ii) MOLECULE TYPE: US28 receptor antisense
 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:
AGCGATGATG CGATAACAAA 20
- 10
(2) INFORMATION FOR SEQ ID NO:8:
 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 20
 (B) TYPE: nucleic acid
15 (C) STRANDEDNESS: single
 (D) TOPOLOGY: unknown
 (ii) MOLECULE TYPE: US28 receptor antisense
 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:
GTCAAATACC ACCACTGGTT 20
- 20
(2) INFORMATION FOR SEQ ID NO:9:
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 (B) TYPE: nucleic acid
25 (C) STRANDEDNESS: single
 (D) TOPOLOGY: unknown
 (ii) MOLECULE TYPE: US27 receptor antisense
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 (B) TYPE: nucleic acid
35 (C) STRANDEDNESS: single
 (D) TOPOLOGY: unknown
 (ii) MOLECULE TYPE: US27 receptor antisense
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 (A) LENGTH: 18
 (B) TYPE: nucleic acid
45 (C) STRANDEDNESS: single
 (D) TOPOLOGY: unknown
 (ii) MOLECULE TYPE: US27 receptor antisense
 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:
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- 50
(2) INFORMATION FOR SEQ ID NO:12:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 18
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
5 (D) TOPOLOGY: unknown
 (ii) MOLECULE TYPE: US27 receptor antisense
 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:
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- 10 (2) INFORMATION FOR SEQ ID NO:13:
 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 18
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
15 (D) TOPOLOGY: unknown
 (ii) MOLECULE TYPE: US27 receptor antisense
 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:
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- 20 (2) INFORMATION FOR SEQ ID NO:14:
 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 18
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
25 (D) TOPOLOGY: unknown
 (ii) MOLECULE TYPE: US27 receptor antisense
 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:
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- 30 (2) INFORMATION FOR SEQ ID NO:15:
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 (A) LENGTH: 18
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
35 (D) TOPOLOGY: unknown
 (ii) MOLECULE TYPE: US27 receptor antisense
 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:
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- 40 (2) INFORMATION FOR SEQ ID NO:16:
 (i) SEQUENCE CHARACTERISTICS:
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 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
45 (D) TOPOLOGY: unknown
 (ii) MOLECULE TYPE: US27 receptor antisense
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- 50 (2) INFORMATION FOR SEQ ID NO:17:
 (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 18
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: unknown
- 5 (ii) MOLECULE TYPE: US27 receptor antisense
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:
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- (2) INFORMATION FOR SEQ ID NO:18:
- 10 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 18
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: unknown
- 15 (ii) MOLECULE TYPE: US27 receptor antisense
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:
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- (2) INFORMATION FOR SEQ ID NO:19:
- 20 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 18
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: unknown
- 25 (ii) MOLECULE TYPE: US27 receptor antisense
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:19:
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- (2) INFORMATION FOR SEQ ID NO:20:
- 30 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 18
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: unknown
- 35 (ii) MOLECULE TYPE: US27 receptor antisense
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:20:
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- (2) INFORMATION FOR SEQ ID NO:21:
- 40 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 18
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: unknown
- 45 (ii) MOLECULE TYPE: US28 receptor antisense
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:21:
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- (2) INFORMATION FOR SEQ ID NO:22:
- 50 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 18

(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: unknown
(ii) MOLECULE TYPE: US28 receptor antisense
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TGATTAAGCA CGTCGGTG 18

(2) INFORMATION FOR SEQ ID NO:23:

(i) SEQUENCE CHARACTERISTICS:
10 (A) LENGTH: 18
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: unknown
(ii) MOLECULE TYPE: US28 receptor antisense
15 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:23:
GAAGAGAAAG ACAACGCC 18

(2) INFORMATION FOR SEQ ID NO:24:

(i) SEQUENCE CHARACTERISTICS:
20 (A) LENGTH: 18
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: unknown
(ii) MOLECULE TYPE: US28 receptor antisense
25 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:24:
GCTGTGGTAC CAGGATAC 18

(2) INFORMATION FOR SEQ ID NO:25:

(i) SEQUENCE CHARACTERISTICS:
30 (A) LENGTH: 18
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: unknown
(ii) MOLECULE TYPE: US28 receptor antisense
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(2) INFORMATION FOR SEQ ID NO:26:

(i) SEQUENCE CHARACTERISTICS:
40 (A) LENGTH: 18
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: unknown
(ii) MOLECULE TYPE: US28 receptor antisense
45 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:26:
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(2) INFORMATION FOR SEQ ID NO:27:

(i) SEQUENCE CHARACTERISTICS:
50 (A) LENGTH: 18
(B) TYPE: nucleic acid

(C) STRANDEDNESS: single
(D) TOPOLOGY: unknown
(ii) MOLECULE TYPE: US28 receptor antisense
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:27:
5 CGGACAGCGT GTCGGAAG 18

(2) INFORMATION FOR SEQ ID NO:28:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 18
10 (B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: unknown
(ii) MOLECULE TYPE: US28 receptor antisense
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:28:
15 GAGACGCGAC ACGCCTCG 18

We claim:

1. An assay for determining therapeutic activity of US28 receptor antagonists, comprising:

(a) obtaining and isolating smooth muscle cells into a first chamber of a migration device, wherein the first migration chamber comprises growth media chambers and is defined by a first side of a membrane and chamber walls, and wherein the migration device comprises a second chamber defined by the second side of the membrane and having an enclosed space;

(b) infecting the smooth muscle cells with human cytomegalovirus (HCVM) containing a gene encoding the US28 receptor;

(c) adding a candidate therapeutic agent to the first chamber; and

(d) determining the amount of cellular migration into the second chamber, whereby inhibition of cellular migration of infected smooth muscle cells indicates therapeutic activity.

2. The assay of claim 1 wherein the smooth muscle cells are isolated from pulmonary arteries.

3. The assay of claim 1 wherein the membrane has a pore size of from about 2 to about 10 microns.

4. The assay of claim 3 wherein the membrane pore size is about 3 microns.

5. The assay of claim 1 wherein the amount of cellular migration is determined by an assay for counting the number of smooth muscle cells in the second chamber wherein the assay for counting the number of smooth muscle cells is selected from the group consisting of microscopic cell counting per unit area, radiolabeling the smooth muscle cells and counting radioactivity in the second chamber, attaching a fluorescent probe to the smooth muscle cells and measuring fluorescence within the second chamber, and combinations thereof.

6. A method for treating atherosclerosis, restenosis, chronic rejection syndrome and graft versus host disease (GVHD), comprising administering an effective amount of an agent that is a US28 receptor antagonist, wherein a US28 receptor antagonist comprises an inhibitor compound that prevents transduction of US28 receptor signal stimulated by a US28 receptor ligand, wherein a US28 receptor ligand is selected from the group consisting of RANTES, MIP-1 α and MCP.

7. The method of claim 6 wherein the US28 receptor antagonist is selected from the group consisting of an antibody that binds to an extracellular portion of the US28 receptor, an antisense oligonucleotide having a nucleic acid sequence antisense to the US28 cDNA and inhibiting translation of US28 expression in infected smooth muscle cells, and the US28 receptor antagonist is selected from the group consisting of an antibody that binds to an extracellular portion of the US28 receptor, and a US28 binding antagonist, wherein the US28 binding antagonist is selected from the group consisting of KHSV encoded vMIP-2, fractalkine, and herbimycin.

8. The method of claim 6 wherein the monoclonal antibody is chimeric or humanized by means for humanizing non-human antibodies.

9. The method of claim 6 wherein the US28 antisense sequences are selected from the group consisting of SEQ ID NOS. 2-28.

5 10. A method for enhancing cellular migration, comprising infecting a cell with a viral nucleic acid containing a gene encoding CVM US28 receptor or transfecting a cell with a vector comprising the cDNA sequence for US28 operably linked to a viral promoter sequence, and stimulating the transfected or infected cell with a US28 receptor ligand, selected from the group consisting of RANTES, MIP-1 α and MCP1.

10

Figure 1.

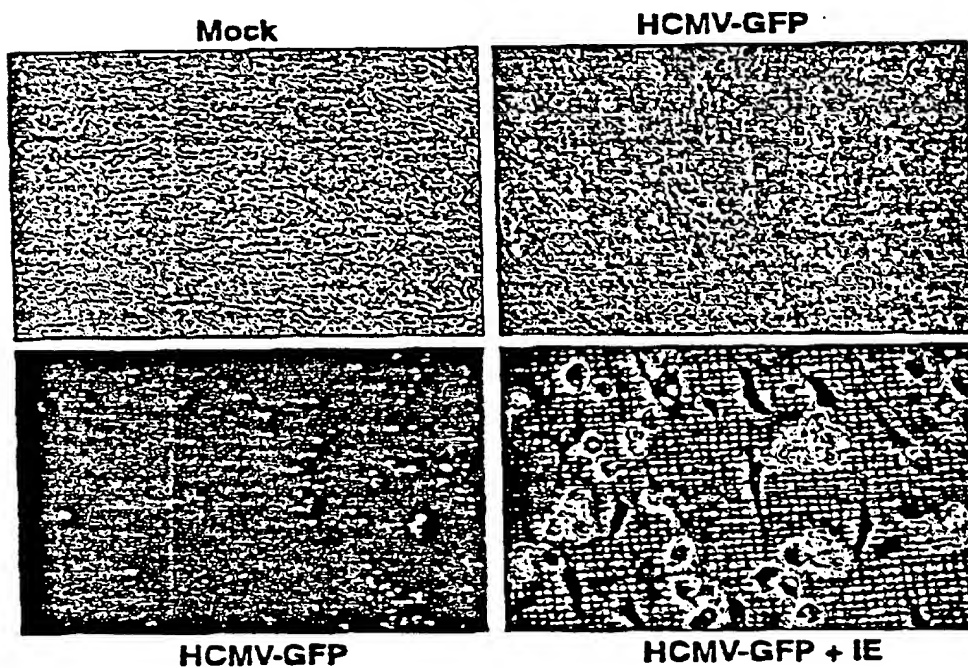
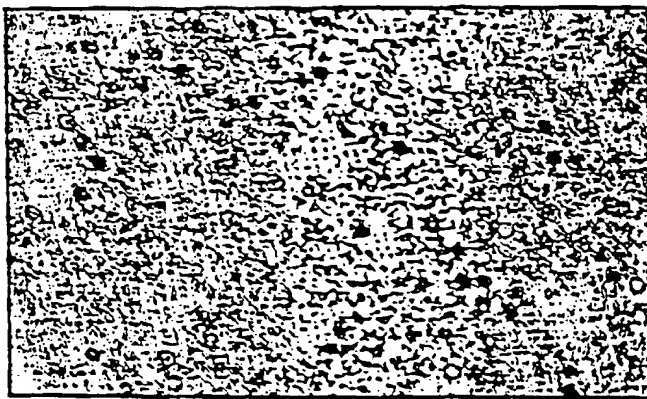
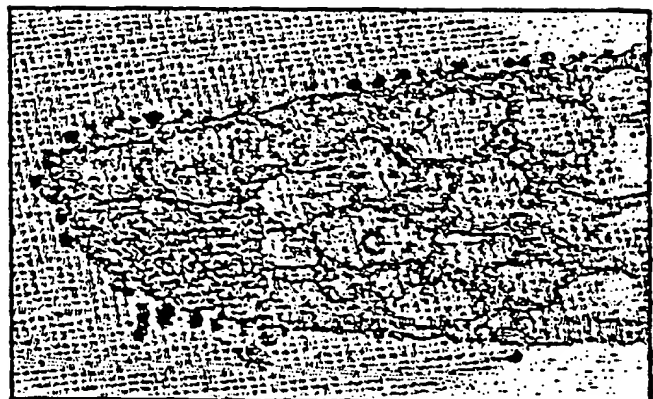
HCMV -GFP Infects SMC

Figure 2.

Em of HCMV Infected SMC



SMC Nucleus



**SMC Cytoplasm and
Plasma Membrane**

Figure 3.

Growth of HCMV in SMC

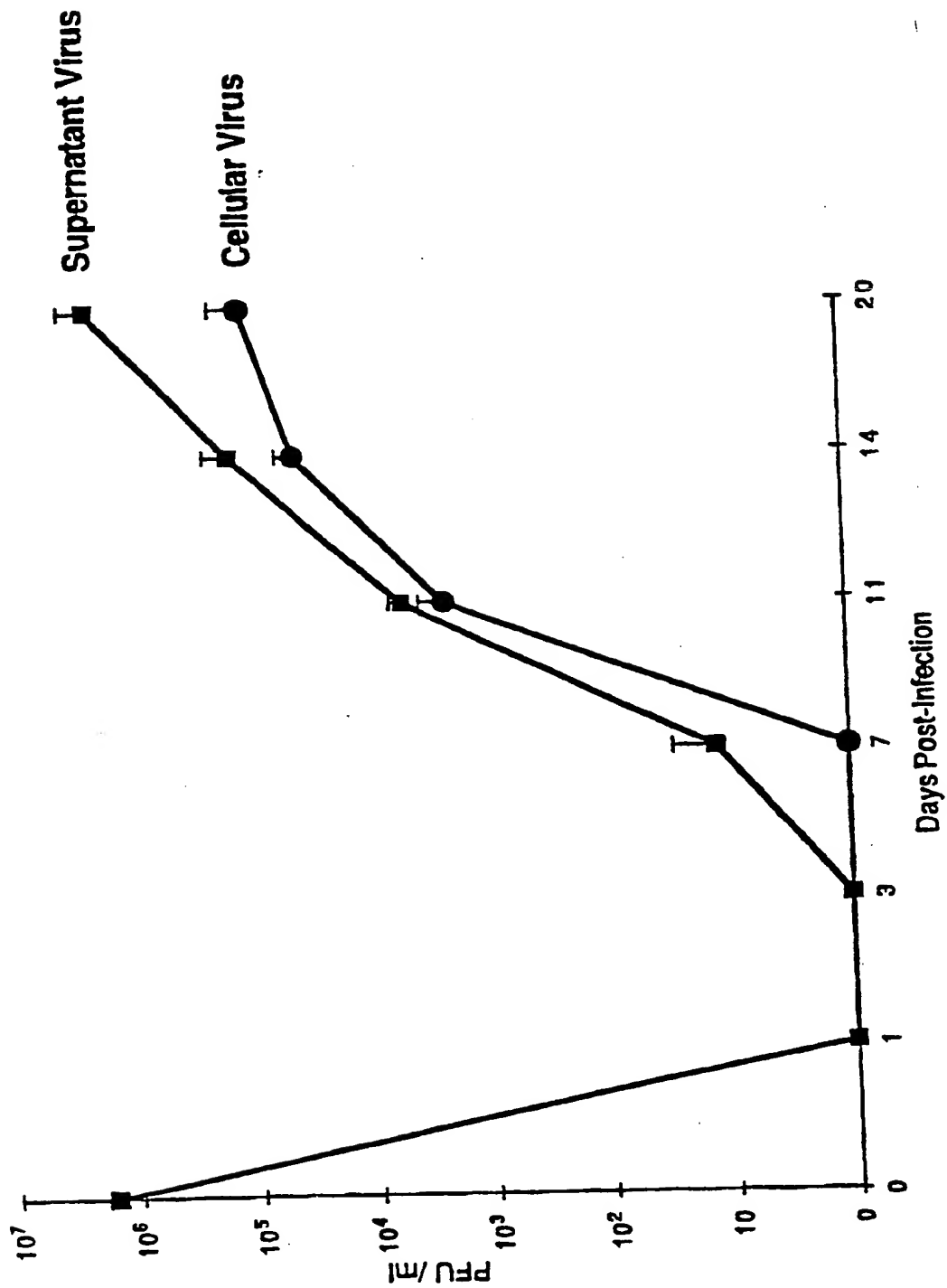


Figure 4.

HCMV Induces Actin Reorganization

Mock

HCMV

PDGF



Figure 5.

Smooth Muscle Cell Migration Assay

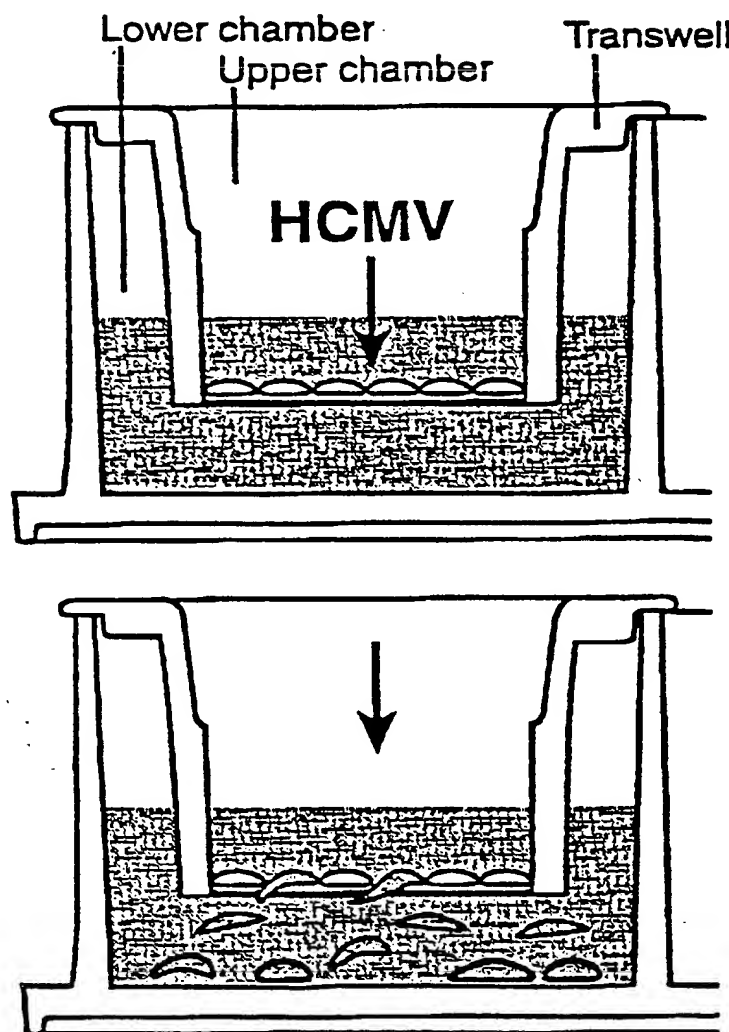


Figure 6.

Presence of HCMV In Migrating SMC

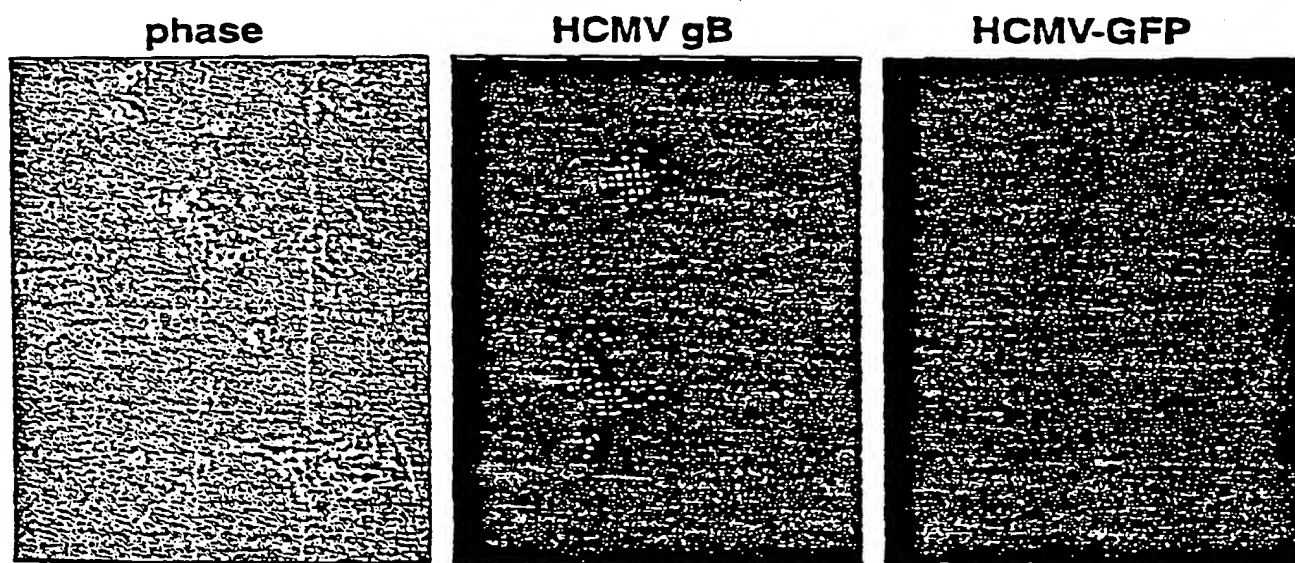


Figure 7.

HCMV Induces Migration of SMC

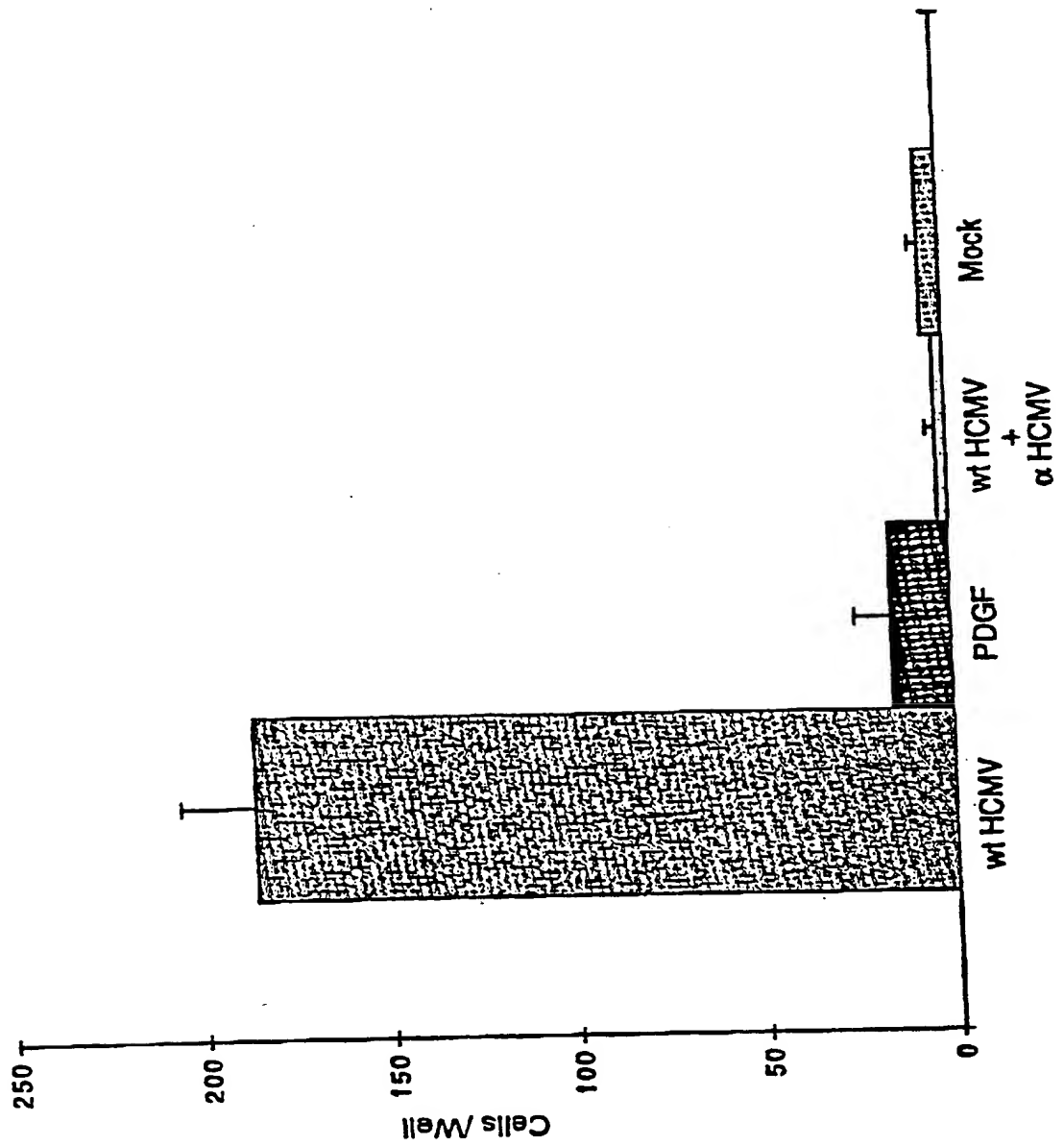


Figure 8.

HCMV Induced Cellular Migration is Unique to SMC

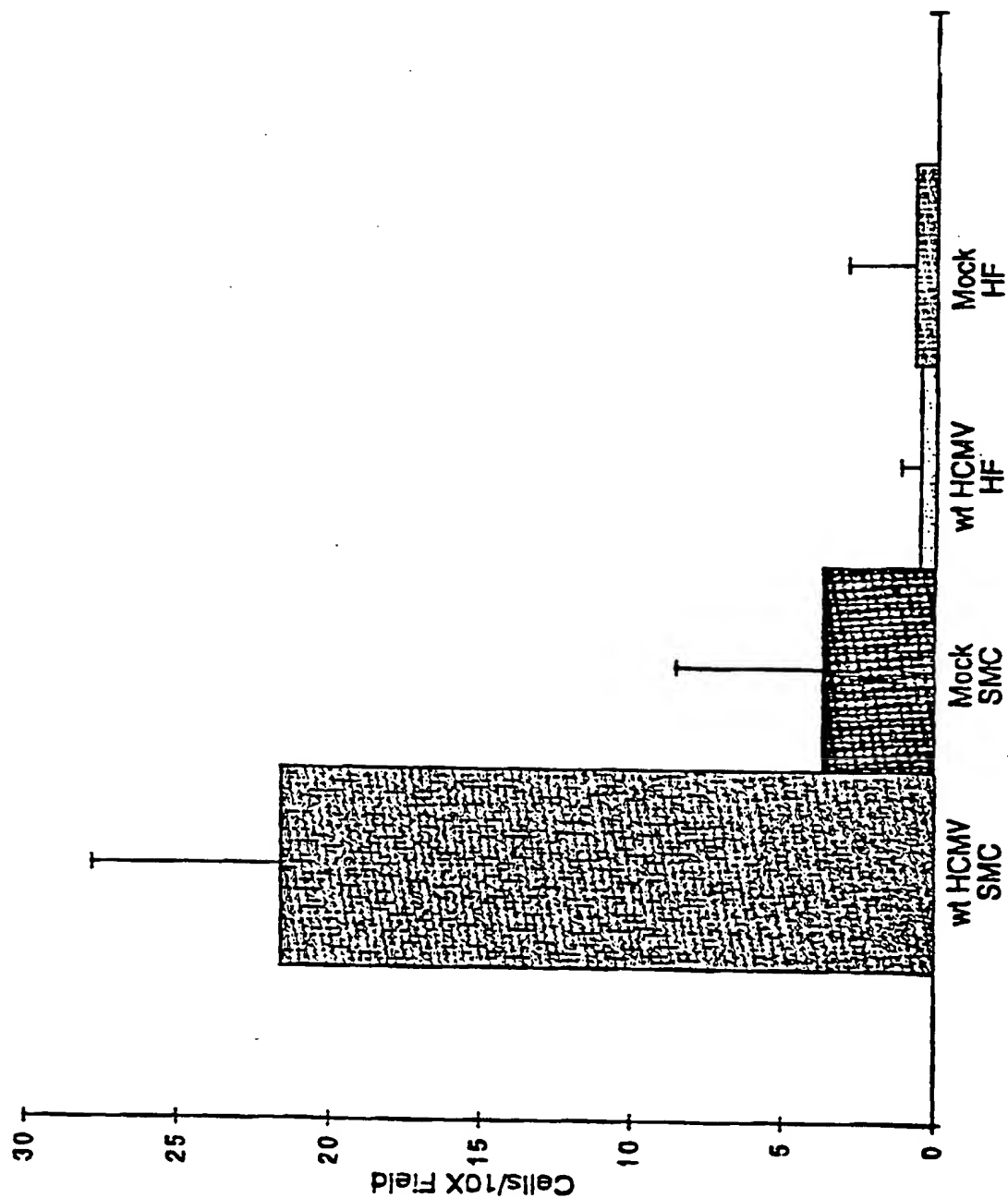


Figure 9.

HCMV Induced SMC Migration Requires De Novo Protein Synthesis

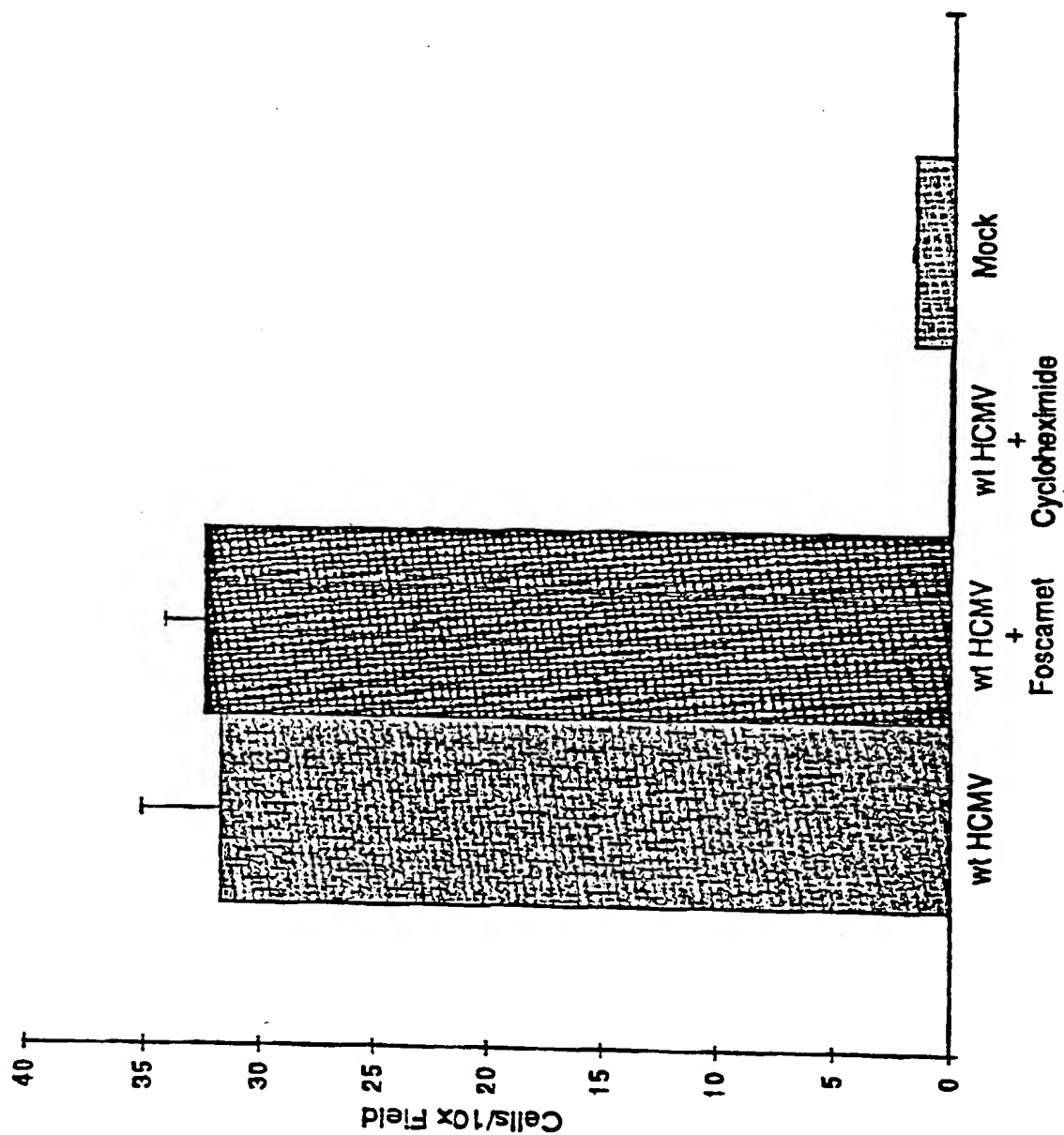


Figure 10.

GCR Related ORF's in the HCMV Genome

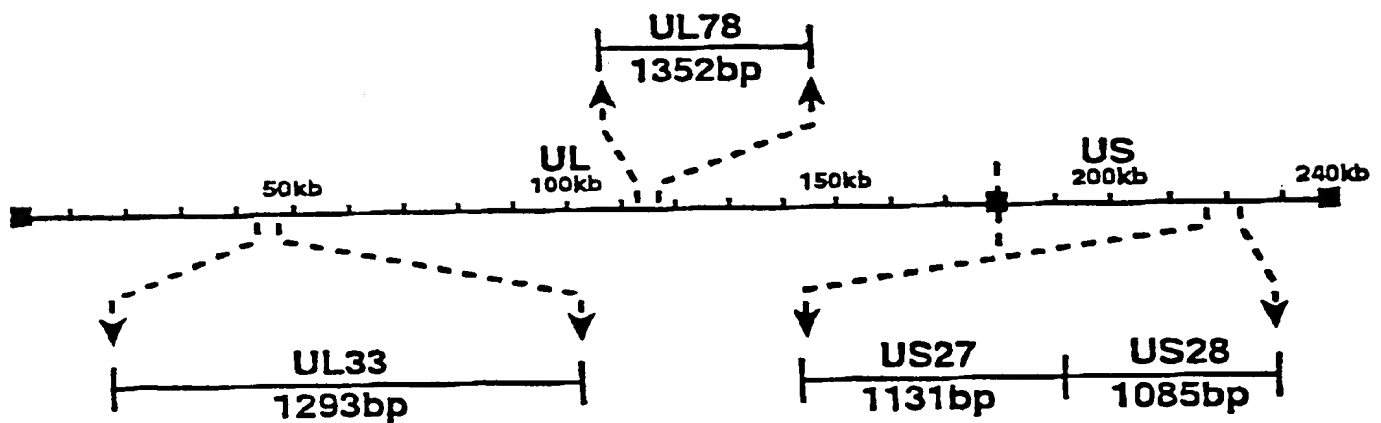


Figure 11.

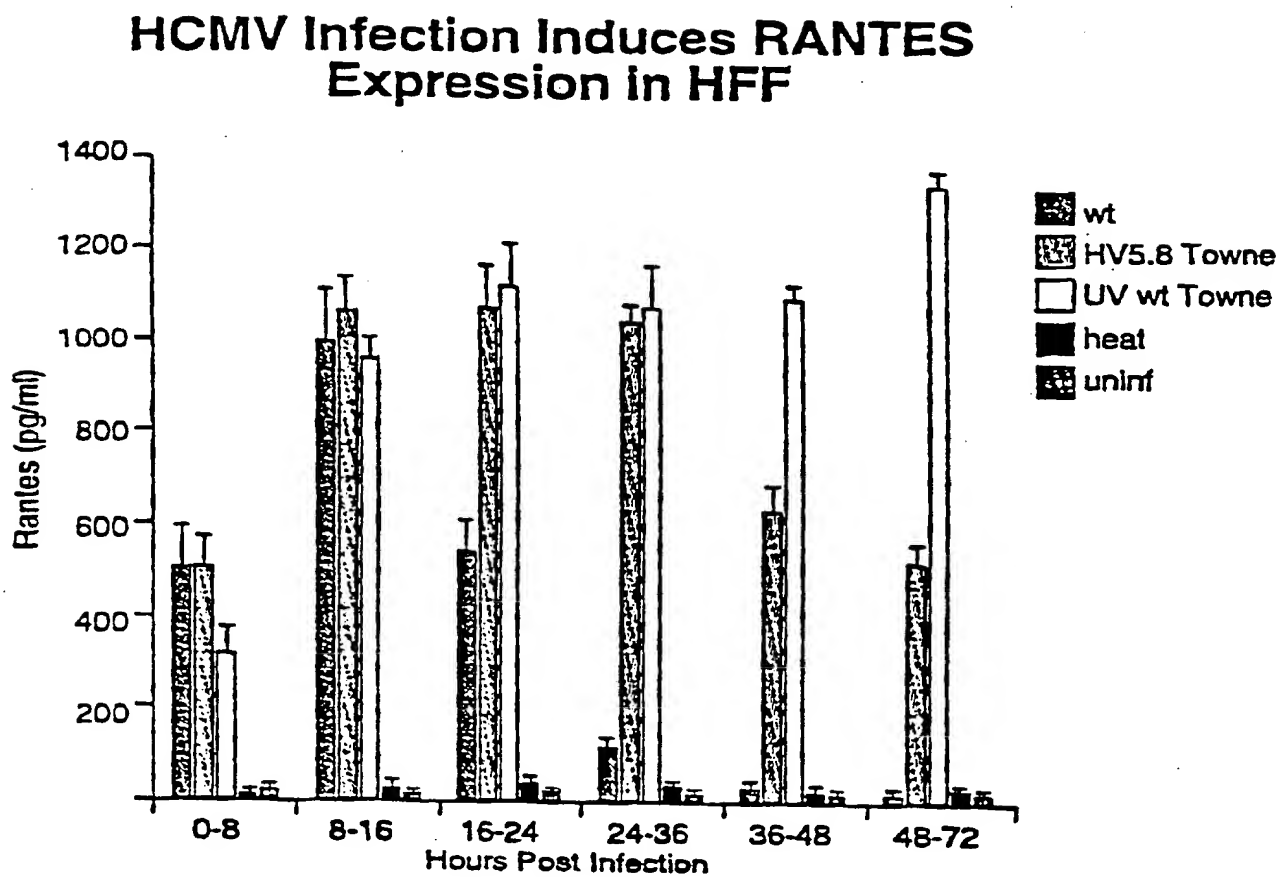
J.Vieira *et al* (J.Virol. in press)

Figure 12.

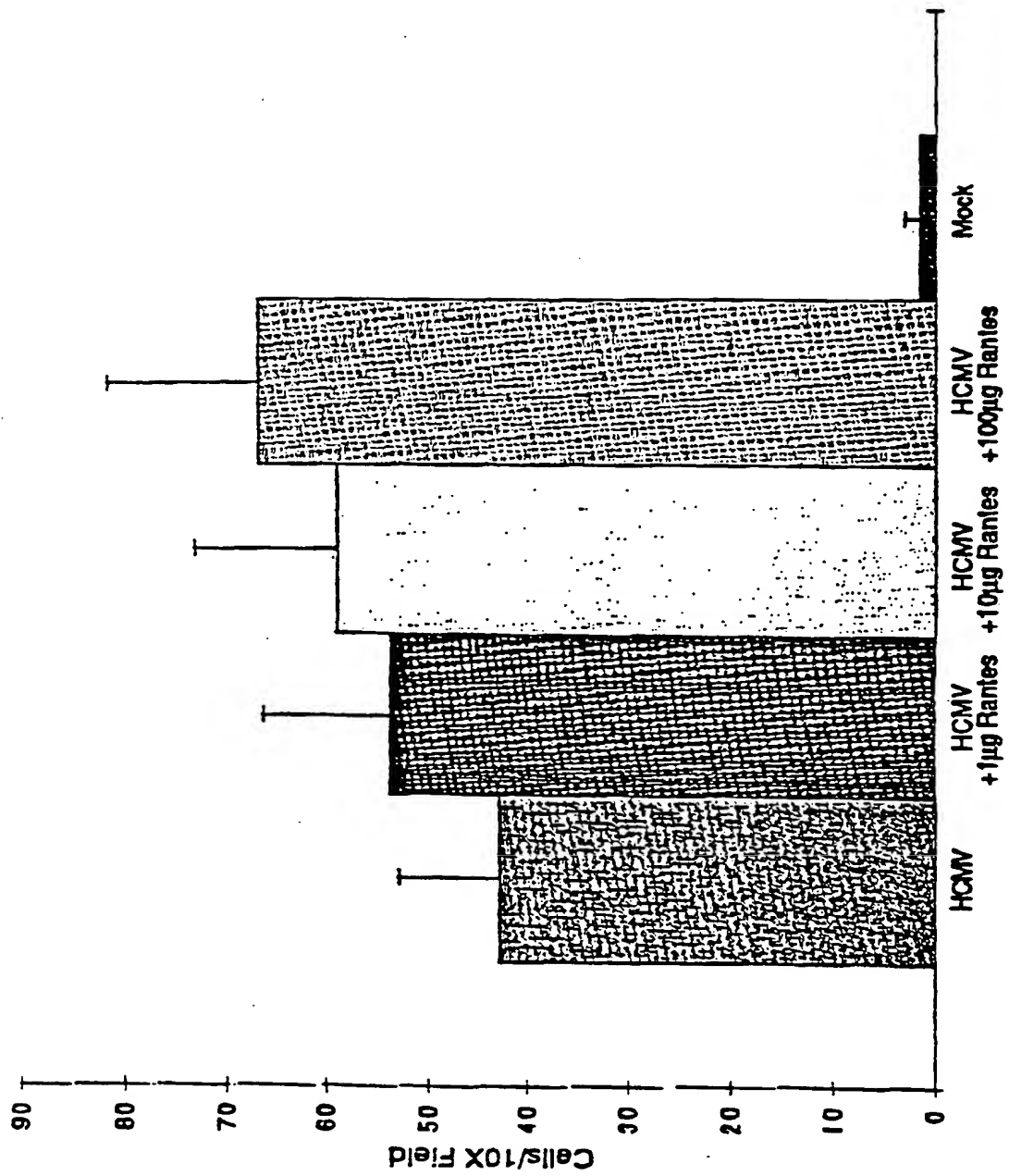
RANTES Ligand Increases SMC Migration

Figure 13.

Deletion of HCMV US28 Inhibits Migration of SMC

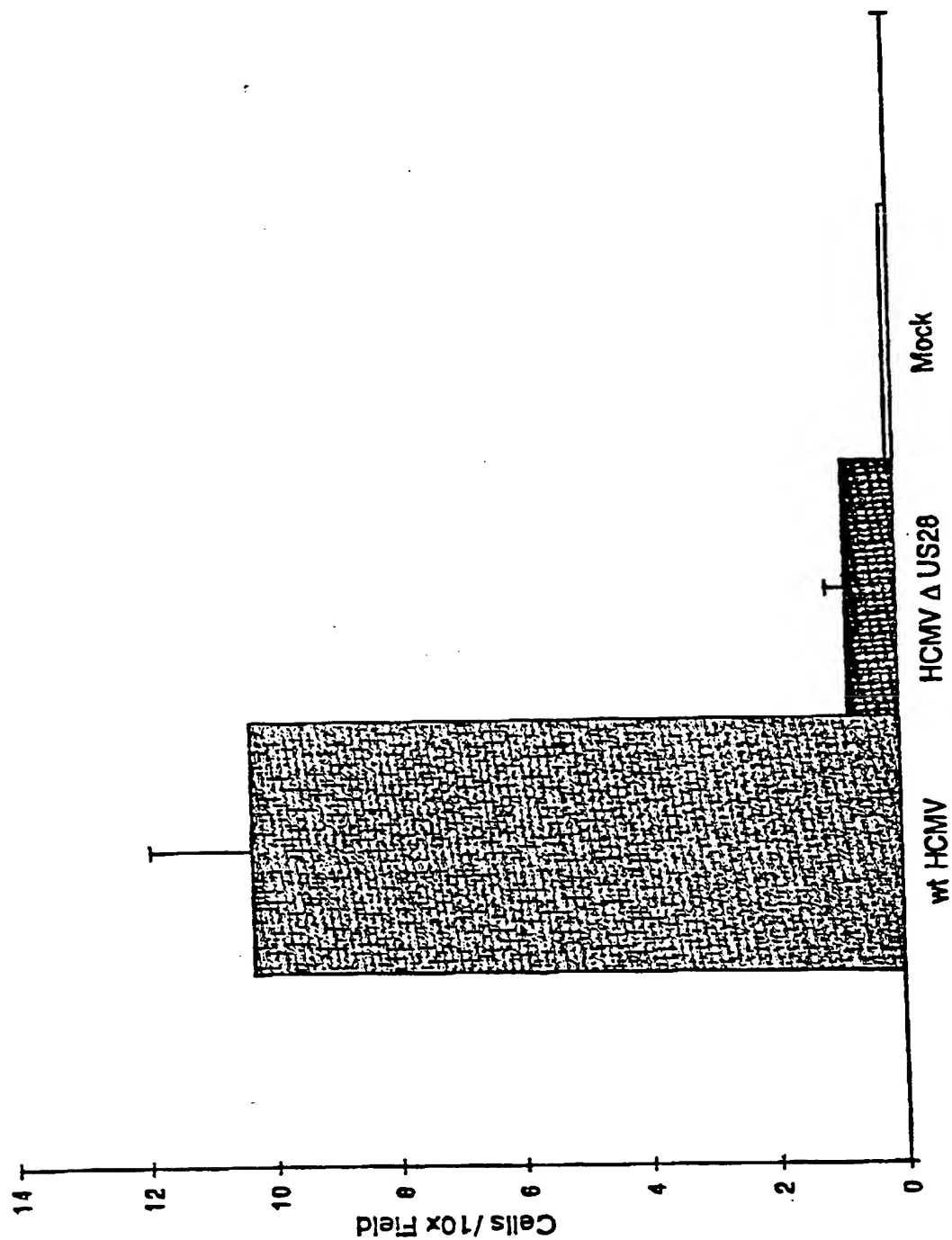


Figure 14.

Construction of HCMV-GFP Recombinants

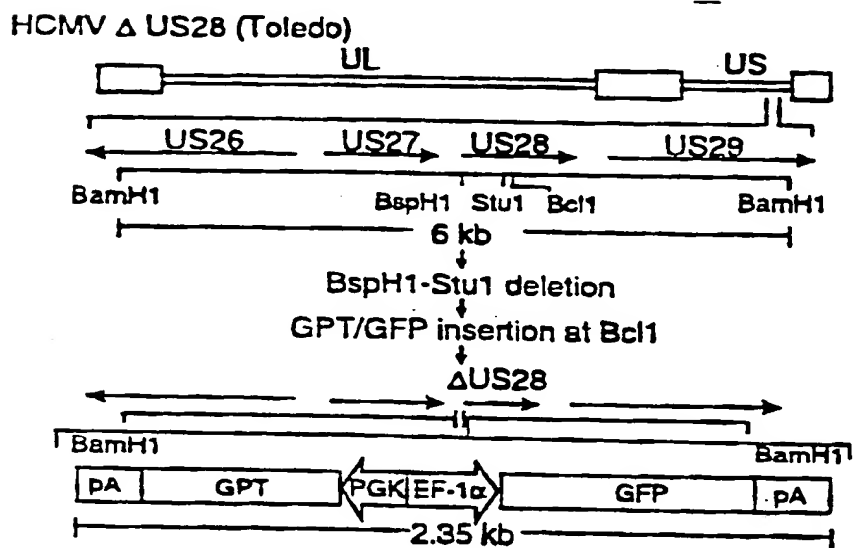
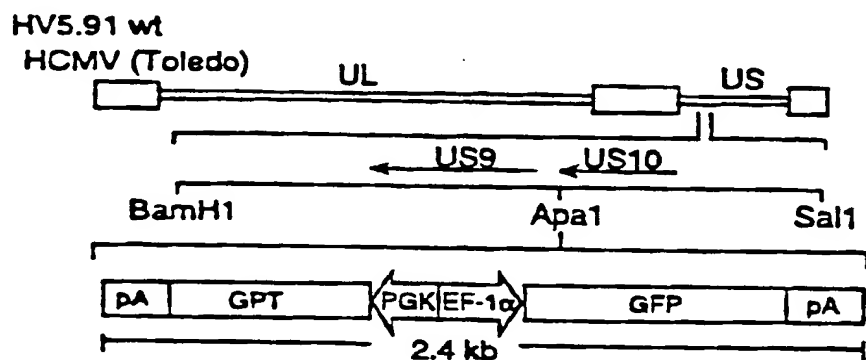
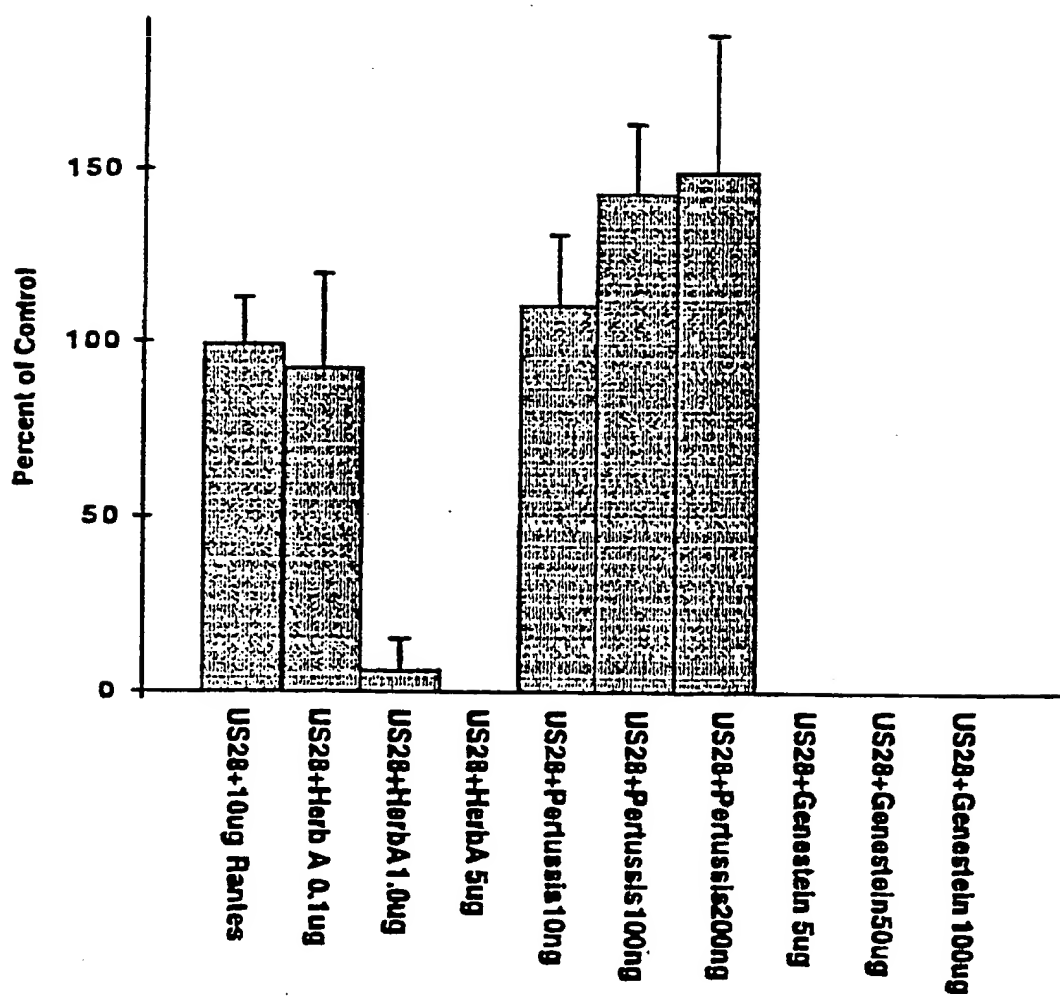


Fig 8



INTERNATIONAL SEARCH REPORT

International application No.
PCT/US99/19990

A. CLASSIFICATION OF SUBJECT MATTER

IPC(7) : Please See Extra Sheet.

US CL : 435/7.21, 69.1; 436/501; 514/2, 41, 324; 530/388.22

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 435/7.21, 69.1; 436/501; 514/2, 41, 324; 530/388.22

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched
NONEElectronic data base consulted during the international search (name of data base and, where practicable, search terms used)
Please See Extra Sheet.

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	BILLSTROM et al. Intracellular Signalling by the Chemokine Receptor US28 during Human Cytomegalovirus Infection. Journal of Virology. July 1998, Vol. 72, No. 7, pages 5535-5544, see entire document.	1-10
A -- X	GAO et al. Human Cytomegalovirus Open Reading Frame US28 Encodes a Functional β Chemokine Receptor. The Journal of Biological Chemistry. 18 November 1994, Vol. 269, No. 46, pages 28539-28542, see especially Figure 3 on page 28541.	1-9 ----- 10



Further documents are listed in the continuation of Box C.



See patent family annex.

* Special categories of cited documents:	*T* later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
A document defining the general state of the art which is not considered to be of particular relevance	*X* document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
E earlier document published on or after the international filing date	*Y* document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
L document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	*G* document member of the same patent family
O document referring to an oral disclosure, use, exhibition or other means	
P document published prior to the international filing date but later than the priority date claimed	

Date of the actual completion of the international search
29 NOVEMBER 1999Date of mailing of the international search report
18 JAN 2000Name and mailing address of the ISA/US
Commissioner of Patents and Trademarks
Box PCT
Washington, D.C. 20231

Facsimile No. (703) 305-3230

Authorized officer

JOHN D. ULM

Telephone No. (703) 308-0196

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US99/19990

C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A ---- Y	HA et al. Atherogenic lipoproteins enhance mesangial cell expression of platelet-derived growth factor; Role of protein tyrosine kinase and cyclic AMP-dependent protein kinase A. Journal of Laboratory Clinical Medicine. May 1998, Vol. 131, No. 5, pages 456-465, see entire document.	1-5, 8-10 ----- 6, 7
A --- Y	KOYAMA et al. Heparan Sulfate Proteoglycans Mediate a Potent Inhibitory Signal for Migration of Vascular Smooth Muscle Cells. Circulation Research. 10 August 1998, Vol. 83, No. 3, pages 305-315. see entire document.	1-5, 8-10 ----- 7, 8

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US99/19990

A. CLASSIFICATION OF SUBJECT MATTER:
IPC (7):

A01N 37/18, 43/04, 43/40; C07K 16/28; C12P 21/06; G01N 33/566, 33/567

B. FIELDS SEARCHED

Electronic data bases consulted (Name of data base and where practicable terms used):

WEST, STN/MEDLINE

search terms: US28, CMV, cytomegalovirus, migration, chamber, herbimycin, atherosclerosis, restenosis, administration, transplant?, fractalkine,

